

Molecular dynamics workshop

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- <https://ub.cbm.uam.es/downloads/MD-workshop-gr-2026.pdf>
- <https://tinyurl.com/MDGRWS26>)

Create Linux VM (if missing)

- Download image: <https://tinyurl.com/LU22BIO>
- Uncompress it
- Launch Virtual Box
 - Click on “Nueva”
 - Nombre: “Bioinfo-Lubuntu22”
 - Tipo: “Linux”
 - Version: “Lubuntu (64-bit)”
 - Click on “Siguiente”
 - Memoria base: 3072
 - Procesadores: 4
 - Click on “Siguiente”
 - “Usar un archivo de disco virtual existente”
 - Path to uncompressed file
 - Click on “Siguiente”
 - Click on “Terminar”
 - Select the new virtual machine and click on “Configuración”
 - Click on “Pantalla” and increase “Memoria de vídeo” up to 128MB

Gromacs install with mamba (skip)

Open terminal



Install micromamba:

- `"${SHELL}" <(curl -L micro.mamba.pm/install.sh)`
 - Exit terminal and open a new one
- micromamba config append channels bioconda --env**
- micromamba update -n base -c defaults conda**
- micromamba activate base**
- micromamba install -c cdat xmgrace**
- micromamba install pocl gromacs vmd**

Prepare environment

Open console:



Activate environment:

```
micromamba activate base
```

Set variables:

```
exampdir=/home/$USER/examp-md-cdk2
```

```
workdir=/home/$USER/md-cdk2
```

```
pdb=cdk2_mod.BL00010001.pdb
```

```
pdbr=cdk2_mod.BL00010001
```

```
export DSSP=~/.bin/dssp
```

Prepare environment II

Make example and working directories:

```
mkdir $workdir
```

```
mkdir $exampdir
```

Go to example files directory:

```
cd $exampdir
```

Get files and uncompress files:

```
wget https://saco.csic.es/s/yeLKeqZmWijWLQS
```

```
tar -xzf cdk2-md.tgz
```

Molecular dynamics simulations

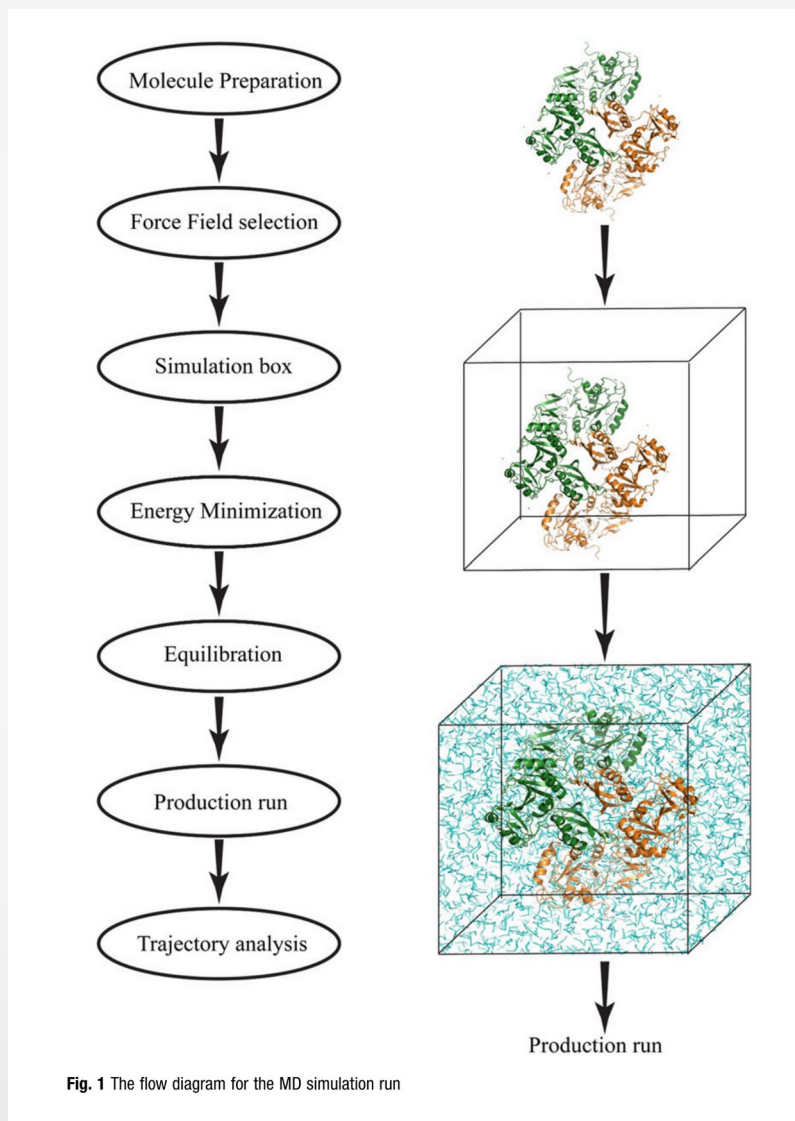


Fig. 1 The flow diagram for the MD simulation run

Redkar, A.S., Ramakrishnan, V. (2023). Molecular Dynamics Simulations. In: Ramakrishnan, V. (eds) Biophysical Characterization of Functional Peptides. Springer Protocols Handbooks. Humana, New York, NY. https://doi.org/10.1007/978-1-0716-3405-9_3

System preparation

- Go to working directory:
cd \$workdir
- copy model structure:
cp \$exampdir/cdk2_mod.BL00010001.pdb .
- copy gromacs run definitions:
cp \$exampdir/*.mdp .
- Add hydrogens and generate topology file:
gmx pdb2gmx -f \$pdb -water tip3p -ff amber03 -ignh
- Generated files:
Topology: topol.top
Coordinates: conf.gro
List of restrained atoms: posre.itp

Water box and ions

- Define water box size and shape:

```
gmx editconf -d 1.0 -bt triclinic -o ${pdbr}_newbox.gro
```

- Add waters to the system:

```
gmx solvate -cp ${pdbr}_newbox.gro -cs spc216.gro -o ${pdbr}_solv.gro -p topol.top
```

- This generates new topology and coordinates files:

```
cdk2_mod.BL00010001_solv.gro  
topol.top
```

- Prepare the system for adding ions

```
gmx grompp -f ions.mdp -c ${pdbr}_solv.gro -p topol.top -o
```

- This generates gromacs input:

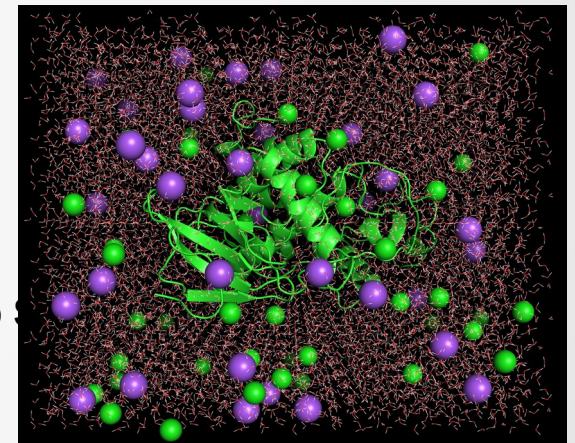
- cdk2_mod.BL00010001_ions.tpr

- Add ions to neutralize protein net charge and to a final concentration of 0.15M

```
echo 13|gmx genion -s ${pdbr}_ions.tpr -o ${pdbr}_ions.gro -p topol.top -neutral -conc 0.15
```

- This generates new topology and coordinates files:

```
cdk2_mod.BL00010001_ions.gro  
topol.top
```



Minimization

- Prepare the system

```
gmx grompp -f minim.mdp -c ${pdbr}_ions.gro -p topol.top -o ${pdbr}_em.tpr
```

- Run the minimization (~ 2min)

```
gmx mdrun -v -deffnm cdk2_mod.BL00010001_em
```

Minimization

- Minimization input file:

; minim.mdp - used as input into grompp to generate em.tpr

; Parameters describing what to do, when to stop and what to save

integrator = steep ;Algorithm (steep = steepest descent minimization)

emtol = 1.0 ; Stop minimization when the maximum force < 1.0 kJ/mol/nm

emstep = 0.01 ; Energy step size

nsteps = 1000 ; Maximum number of (minimization) steps to perform

; Parameters describing how to find the neighbors of each atom and how to calculate the interactions

nstlist = 10 ; Frequency to update the neighbor list and long range forces

ns_type = grid ; Method to determine neighbor list (simple, grid)

rlist = 1.0 ; Cut-off for making neighbor list (short range forces)

coulombtype = PME ; Treatment of long range electrostatic interactions

rcoulomb = 1.0 ; Short-range electrostatic cut-off

rvdw = 1.0 ; Short-range Van der Waals cut-off

pcbc = xyz ; Periodic Boundary Conditions (yes/no)

-

Minimization analysis

- Generated files

```
cdk2_mod.BL00010001_em.edr #Energies
```

```
cdk2_mod.BL00010001_em.trr #Trajectory
```

```
cdk2_mod.BL00010001_em.log #Log
```

```
cdk2_mod.BL00010001_em.gro #Final coordinates
```

- Analyze potential energy evolution

```
echo '10 0' | gmx energy -f ${pdbr}_em.edr -o ${pdbr}_potential.xvg
```

- View graph

```
xmgrace ${pdbr}_potential.xvg
```

Equilibration NVT with restrains

- NVT: Canonical ensemble (amount, volume and temperature constant)
- Prepare the system

```
gmx grompp -f nvt.mdp -c ${pdbr}_em.gro -r ${pdbr}_em.gro -p  
topol.top -o ${pdbr}_nvt.tpr
```

- Run the equilibration (5ps, ~ 1min)

```
gmx mdrun -deffnm ${pdbr}_nvt
```

Equilibration NVT Input file

- title = Amber03 NVT equilibration
- **define = -DPOSRES** ; position restrain the protein
- ; Run parameters
- integrator = md ; leap-frog integrator
- **nsteps = 2500** ; 2fs * 2500steps = 5 ps
- dt = 0.002 ; 2 fs
- ; Output control
- ;nstxout = 100 ; save coordinates every 0.2 ps
- ;nstvout = 100 ; save velocities every 0.2 ps
- nstxout-compressed = 100 ; xtc compressed trajectory output every 0.2ps
- nstenergy = 100 ; save energies every 0.2 ps
- nstlog = 100 ; update log file every 0.2 ps
- nstcalcenergy = -1
- ; Bond parameters
- continuation = **no** ; first dynamics run
- constraint_algorithm = lincs ; holonomic constraints
- constraints = h-bonds ; bonds involving H are constrained
- lincs_iter = 1 ; accuracy of LINCS
- lincs_order = 4 ; also related to accuracy
- ; Neighborsearching
- cutoff-scheme = Verlet ; Buffered neighbor searching
- ns_type = grid ; search neighboring grid cells
- nstlist = 10 ; 20 fs, largely irrelevant with Verlet scheme
- rlist = 1.0 ; short-range neighborlist cutoff (in nm)
- rcoulomb = 1.0 ; short-range electrostatic cutoff (in nm)
- rvdw = 1.0 ; short-range van der Waals cutoff (in nm)
- ; Electrostatics
- coulombtype = PME ; Particle Mesh Ewald for long-range electrostatics
- pme_order = 4 ; cubic interpolation
- fourierspacing = 0.12 ; grid spacing for FFT
- ; Temperature coupling is on
- tcoupl = V-rescale ; modified Berendsen thermostat
- tc-grps = Protein Non-Protein ; two coupling groups - more accurate
- tau_t = 0.1 0.1 ; time constant, in ps
- ref_t = 300 300 ; reference temperature, one for each group, in K
- ; Pressure coupling is off
- **pcoupl = no** ; no pressure coupling in NVT
- ; Periodic boundary conditions
- pbc = xyz ; 3-D PBC
- ; Dispersion correction
- DispCorr = EnerPres ; account for cut-off vdW scheme
- ; Velocity generation
- **gen_vel = yes** ; assign velocities from Maxwell distribution
- **gen_temp = 300** ; temperature for Maxwell distribution
- gen_seed = -1 ; generate a random seed

Equilibration NVT analysis

- Generated files

cdk2_mod.BL00010001_nvt.cpt #Checkpoint

cdk2_mod.BL00010001_nvt.edr #Energies

cdk2_mod.BL00010001_nvt.xtc #Trajectory

cdk2_mod.BL00010001_nvt.log #Log

cdk2_mod.BL00010001_nvt.gro #Final coordinates

- Check temperature evolution

```
echo '16 0' | gmx energy -f ${pdbr}_nvt.edr -o ${pdbr}_temperature.xvg
```

- View graph

```
xmgrace ${pdbr}_temperature.xvg
```

Equilibration NPT with restrains

- Prepare the system

```
gmx grompp -f npt.mdp -c ${pdbr}_nvt.gro -r ${pdbr}_nvt.gro -t ${pdbr}_nvt.cpt -p  
topol.top -o ${pdbr}_npt.tpr
```

- Run the equilibration (15 ps, ~ 3min)

```
gmx mdrun -deffnm ${pdbr}_npt
```

Equilibration NPT Input file

```

title                = Amber03 NPT equilibration
define               = -DPOSRES ; position restrain the
protein
; Run parameters
Integrator           = md          ; leap-frog integrator
Nsteps               = 7500        ; 2fs * 7500steps = 15ps
Dt                   = 0.002      ; 2 fs
; Output control
;nstxout              = 100        ; save coordinates every 0.2 ps
;nstvout              = 100        ; save velocities every 0.2 ps
Nstxout-compressed  = 100         ; xtc compressed trajectory
output every 0.2 ps
nstenergy             = 500
; save energies every 1 ps
nstlog                = 100
; update log file every 0.2 ps
; Bond parameters
Continuation       = yes
; Restarting after NVT
constraint_algorithm = lincs      ; holonomic constraints
Constraints           = h-bonds   ; bonds involving H are
constrained
lincs_iter            = 1         ; accuracy of LINCS
lincs_order           = 4         ; also related to accuracy
; Neighborsearching
ns_type               = grid
; search neighboring grid cells
Nstlist               = 10        ;20 fs, largely irrelevant with
Verlet scheme
cutoff-scheme         = Verlet
Rlist                 = 1.0       ; short-range neighborlist cutoff
(in nm)
Rcoulomb              = 1.0       ; short-range electrostatic
cutoff (in nm)
Rvdw                  = 1.0       ; short-range van der Waals
cutoff (in nm)
; Electrostatics
coulombtype           = PME       ; Particle Mesh Ewald
for long-range electrostatics
pme_order             = 4         ; cubic interpolation
Fourierspacing        = 0.12     ; grid spacing for FFT
; Temperature coupling is on
tcoupl                = V-rescale ; modified Berendsen
thermostat
tc-grps               = Protein Non-Protein ; two
coupling groups - more accurate
tau_t                 = 0.1 0.1   ; time constant, in ps
ref_t                  = 300 300   ; reference
temperature, one for each group, in K
; Pressure coupling is on
pcoupl                = Berendsen ; Pressure coupling
on in NPT
Pcoupltype            = isotropic ; uniform scaling of
box vectors
tau_p                  = 2.0       ; time constant, in
ps
ref_p                  = 1.0       ; reference
pressure, in bar
compressibility        = 4.5e-5    ; isothermal
compressibility of water, bar-1
refcoord_scaling      = com
; Periodic boundary conditions
pbc                   = xyz       ; 3-D PBC
; Dispersion correction
DispCorr              = EnerPres  ; account for cut-off
vdW scheme
; Velocity generation
gen_vel               = no        ; Velocity
generation is off

```

Equilibration NPT analysis

- Check pressure evolution

```
echo '18 0' | gmx energy -f ${pdbr}_npt.edr -o ${pdbr}_pressure.xvg
```

- View graph

```
xmgrace cdk2_mod.BL00010001_pressure.xvg
```

- Check density evolution

```
echo '24 0' | gmx energy -f ${pdbr}_npt.edr -o ${pdbr}_density.xvg
```

- View graph

```
xmgrace cdk2_mod.BL00010001_density.xvg
```

- Analyze rmsd evolution

```
echo '4 4' | gmx rms -s ${pdbr}_npt.tpr -f ${pdbr}_npt.xtc -o ${pdbr}-bkbone-rmsd-npt.xvg
```

- View graph

```
xmgrace cdk2_mod.BL00010001-bkbone-rmsd-npt.xvg
```

MD production

- Prepare the system

```
gmx grompp -f md.mdp -c ${pdbr}_npt.gro -t ${pdbr}_npt.cpt -p topol.top -o ${pdbr}_md.tpr
```

- Run the production (40ps, 5min)

```
gmx mdrun -deffnm ${pdbr}_md -cpt 5
```

MD production Input file

```

title = Amber03 MD
; Don't use restrains
; Run parameters
Integrator = md ; leap-frog integrator
Nsteps = 20000 ; 2 * 20000 = 40 ps
Dt = 0.002 ; 2 fs
; Output control
;nstxout = 100 ; save coordinates every 0.2 ps
;nstvout = 100 ; save velocities every 0.2 ps
Nstxout-compressed = 100 ; xtc compressed trajectory
output every 2 ps
nstenergy = 500 ;
save energies every 1 ps
nstlog = 500 ;
update log file every 1 ps
; Bond parameters
continuation = yes ; Restarting
after NPT
constraint_algorithm = lincs ; holonomic constraints
constraints = h-bonds ; bonds involving H are
constrained
lincs_iter = 1 ; accuracy of LINCS
lincs_order = 4 ; also related to accuracy
; Neighborsearching
cutoff-scheme = Verlet ; Buffered neighbor
searching
ns_type = grid ; search neighboring grid
cells
Nstlist = 10 ; 20 fs, largely irrelevant with
Verlet scheme
Rcoulomb = 1.0 ; short-range electrostatic
cutoff (in nm)
Rvdw = 1.0 ; short-range van der Waals
cutoff (in nm)
; Electrostatics
Coulombtype = PME ; Particle Mesh
Ewald for long-range electrostatics
pme_order = 4
; cubic interpolation
Fourierspacing = 0.12 ; grid spacing for
FFT
; Temperature coupling is on
tcoupl = V-rescale ; modified
Berendsen thermostat
tc-grps = Protein Non-Protein ; two coupling
groups - more accurate
tau_t = 0.1 0.1 ; time constant,
in ps
ref_t = 300 300 ; reference
temperature, one for each group, in K
; Pressure coupling is on
pcoupl = Parrinello-Rahman ; Pressure
coupling on in NPT
Pcoupltype = isotropic ; uniform scaling
of box vectors
tau_p = 2.0 ; time constant,
in ps
ref_p = 1.0 ; reference
pressure, in bar
compressibility = 4.5e-5 ; isothermal
compressibility of water, bar^-1
; Periodic boundary conditions
pbc = xyz ; 3-D PBC
; Dispersion correction
DispCorr = EnerPres ; account for cut-
off vdW scheme
; Velocity generation
gen_vel = no ; Velocity
generation is off

```

MD production analysis I

- Check energy evolution

```
echo '13 0' | gmx energy -f ${pdbr}_md.edr -o ${pdbr}_ener_md.xvg
```

View graph

```
xmgrace ${pdbr}_ener_md.xvg
```

- Check temperature evolution

```
echo '15 0' | gmx energy -f ${pdbr}_md.edr -o ${pdbr}_temp_md.xvg
```

- View graph

```
xmgrace ${pdbr}_temp_md.xvg
```

- Check pressure evolution

```
echo '17 0' | gmx energy -f ${pdbr}_md.edr -o ${pdbr}_press_md.xvg
```

- View graph

```
xmgrace ${pdbr}_press_md.xvg
```

- Remove waters and ions from trajectory

```
echo '1 1' | gmx trjconv -pbc mol -center -s ${pdbr}_md.tpr -f ${pdbr}_md.xtc -o ${pdbr}_md_nw.xtc
```

- Generate parameters file without water and ions

```
echo '1' | gmx convert-tpr -s ${pdbr}_md.tpr -o ${pdbr}_md_nw.tpr
```

- Extract first frame from production to .gro

```
echo '0' | gmx trjconv -f ${pdbr}_md_nw.xtc -s ${pdbr}_md_nw.tpr -o ${pdbr}_md_nw.gro -dump 0
```

- View trajectory

```
vmd ${pdbr}_md_nw.gro -xtc ${pdbr}_md_nw.xtc
```

MD production analysis II

- Analyze fluctuations by residue

```
echo 1| gmx rmsf -s ${pdbr}_md_nw.tpr -f ${pdbr}_md_nw.xtc -o ${pdbr}-  
rmsf-md.xvg -ox ${pdbr}-md-avg.pdb -res -oq ${pdbr}-md-bfact.pdb
```

- View graph

```
xmgrace ${pdbr}-rmsf-md.xvg
```

- View PDB

```
pymol ${pdbr}-md-bfact.pdb
```

Action → Preset → b factor putty

- Analyze RMSD

```
echo '4 4' |gmx rms -s ${pdbr}_md_nw.tpr -f ${pdbr}_md_nw.xtc -o ${pdbr}-  
bkbone-rmsd-md.xvg
```

- View graph

```
xmgrace ${pdbr}-bkbone-rmsd-md.xvg
```

Compare with real cdk2 structure

- Set variables:

```
readdir=/home/usuario/3qqg-md
```

```
pdbr=3qqg
```

- Make example directory:

```
cd
```

```
mkdir $readdir
```

- Go to example files directory:

```
cd $readdir
```

- Get files:

```
wget ub.cbm.uam.es/members/dabia/3qqg-md.tgz
```

- Uncompress files:

```
tar -xzf 3qqg-md.tgz
```